PrimeSurface® Publication List

(Updated April 2020) Total >306

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S-BIO – 20 EXECUTIVE DRIVE – HUDSON, NH 03051, USA
TEL: +1 603 546 6821 | FAX: +1 603 577 9968

Publications from 2019 to Apr. 2020 (80)

PrimeSurface® Stem Cell Differentiation Research Publications (41)

< Retinal Research and Eye disease >

1. K. Eastlake, et al, Phenotypic and Functional Characterization of Müller Glia Isolated from Induced Pluripotent Stem Cell-Derived Retinal Organoids: Improvement of Retinal Ganglion Cell Function upon Transplantation. *Stem Cells Translational Medicine*, 2019, 8.8: 775–784 [MS-9096V]
https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6646702/

2. S. Kitahata, et al, Critical Functionality Effects from Storage Temperature on Human Induced Pluripotent Stem Cell-Derived Retinal Pigment Epithelium Cell Suspensions. *Scientific Reports*, 2019, 9:2891 [MS-9096U, M or V]
https://www.nature.com/articles/s41598-018-38065-6

3. C. B. Mellough, et al, Systematic Comparison of Retinal Organoid Differentiation from Human Pluripotent Stem Cells Reveals Stage Specific, Cell Line, and Methodological Differences. *Stem Cells Translational Medicine*, 2019, 8.7: 694–706 [MS-9096V]
https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6591558/

4. H. C. Suen, et al, Transplantation of Retinal Ganglion Cells Derived from Male Germline Stem Cell as a Potential Treatment to Glaucoma. *Stem Cells and Development*, 2019, 28.20:1365-1375 [MS-9096U]
https://pubmed.ncbi.nlm.nih.gov/31580778/

5. A. Kuwahara, et al, Preconditioning the Initial State of Feeder-free Human Pluripotent Stem Cells Promotes Self-formation of Three-dimensional Retinal Tissue. *Scientific Reports*, 2019, 9: 18936. [MS-9096V]
https://www.nature.com/articles/s41598-019-55130-w

< Neuroscience Research >

1. H. Takeuch, et al, A scaffold-free Bio 3D nerve conduit for repair of a 10-mm peripheral nerve defect in the rats. *Microsurgery*, 2020, 40.2: 207-216. [MS-9096U, M or V]
https://onlinelibrary.wiley.com/doi/abs/10.1002/micr.30533

2. S. Mitsuzawa, et al, The efficacy of a scaffold-free Bio 3D conduit developed from autologous dermal fibroblasts on peripheral nerve regeneration in a canine ulnar nerve injury model: a preclinical proof-of-concept study. *Cell transplantation*, 2019, 28.9-10: 1231-1241. [MS-9096U, M or V]
https://journals.sagepub.com/doi/full/10.1177/0963689719855346

3. D. Lukmanto, et al, Dynamic changes of mouse embryonic stem cell-derived neural stem cells under in vitro prolonged culture and hypoxic conditions. *Stem cells and development*, 2019, 28.21: 1434-1450. [MS-9096U, M or V]
https://www.liebertpub.com/doi/abs/10.1089/scd.2019.0101

4. T. Sunohara, et al, MicroRNA-based separation of cortico-fugal projection neuron-like cells derived from embryonic stem cells. *Frontiers in neuroscience*, 2019, 13: 1141. [MS-9096U]

https://www.frontiersin.org/articles/10.3389/fnins.2019.01141/full

5. H. Sakaguchi, et al, Self-organized synchronous calcium transients in a cultured human neural network derived from cerebral organoids. *Stem Cell Reports*, 2019, 13.3: 458-473. [MS-9096V]
https://www.sciencedirect.com/science/article/pii/S2213671119301973

6. W. Zhu, et al, Precisely controlling endogenous protein dosage in hPSCs and derivatives to model FOXG1 syndrome. *Nature communications*, 2019, 10.1: 1-18. [MS-9096VZ]
https://www.nature.com/articles/s41467-019-08841-7

7. S. N. Sarma, et al, Tyroxine hydroxylase-positive neuronal cell population is increased by temporal dioxin exposure at early stage of differentiation from human embryonic stem cells. *International journal of molecular sciences*, 2019, 20.11: 2687. [MS-9096U]
https://www.mdpi.com/1422-0067/20/11/2687/htm?__cf_chl_managed_tk__=kD2hplaZCj1mntbu9quEZZbhq.fr0pkTEHaB7DK7hSc-1641534247-0-gaNycGzNB-U

8. A. Hirota, et al, The nucleosome remodeling and deacetylase complex protein CHD4 regulates neural differentiation of mouse embryonic stem cells by down-regulating p53. *Journal of Biological Chemistry*, 2019, 294.1: 195-209. [MS-9096U, M or V]
https://www.jbc.org/content/294/1/195.full.pdf

9. B, Samata, et al, L1CAM is a marker for enriching corticospinal motor neurons in the developing brain. *Frontiers in cellular neuroscience*, 2020, 14: 31. [MS-9096U, M or V]
https://www.frontiersin.org/articles/10.3389/fncel.2020.00031/full

< Cardiomyocytes and Heart Research >

1. T. Kitsuka, et al, 2-Cl-C.OXT-A stimulates contraction through the suppression of phosphodiesterase activity in human induced pluripotent stem cell derived cardiac organoids. *PLoS ONE,* 2019, 14.7: e0213114 [MS-9096V]
https://journals.plos.org/plosone/article/file?type=printable&id=10.1371/journal.pone.0213114

2. S. Yoshida, et al, Hydrogel Microchambers Integrated with Organic Electrodes for Efficient Electrical Stimulation of Human iPSC-Derived Cardiomyocytes. *Macromolecular Bioscience,* 2020, 19.6: e1900060 [MS-9096U]
https://onlinelibrary.wiley.com/doi/abs/10.1002/mabi.201900060

3. M. Kimura, et al, Increased mesodermal and mesendodermal populations by BMP4 treatment facilitates human iPSC line differentiation into a cardiac lineage. *Journal of Stem Cells & Regenerative Medicine,* 2019, 15.2: 45-51 [MS-90900]
https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6971377/

< Bone and Cartilage Research >

1. K. Endo, et al, Effect of Fibroblast Growth Factor-2 and Serum on Canine Mesenchymal Stem Cell Chondrogenesis. *Tissue Engineering Part A,* 2019, 25: 11-12 [MS-9096U]

https://www.liebertpub.com/doi/10.1089/ten.TEA.2018.0177?url_ver=Z39.88-2003&rfr_id=ori%3Arid%3Acrossref.org&rfr_dat=cr_pub++0pubmed

2. T. Oshima, et al, A Scaffold-Free Allogeneic Construct From Adipose-Derived Stem Cells Regenerates an Osteochondral Defect in a Rabbit Model. *Arthroscopy: The Journal of Arthroscopic & Related Surgery,* 2019, 35.2: 583-593 [MS-9096U]
https://www.sciencedirect.com/science/article/abs/pii/S0749806318307242

3. K. Endo, et al, Comparison of the effect of growth factors on chondrogenesis of canine mesenchymal stem cells. *The Journal of Veterinary Medical Science,* 2019, 81.8: 1211-1218 [MS-9096U, M or V]
https://www.jstage.jst.go.jp/article/jvms/advpub/0/advpub_18-0551/_pdf/-char/ja

4. A. Yamasaki, et al, Osteochondral regeneration using constructs of mesenchymal stem cells made by bio three-dimensional printing in mini-pigs. *Journal of Orthopaedic Research,* 2019, 37.6: 1398-1408 [MS-9096U]
https://onlinelibrary.wiley.com/doi/abs/10.1002/jor.24206

5. E. K. Breathwaite, et al, Scaffold-free bioprinted osteogenic and chondrogenic systems to model osteochondral physiology. *Biomedical Materials,* 2019, 14.6: 65010 [MS-9096U]
https://iopscience.iop.org/article/10.1088/1748-605X/ab4243/meta

6. S. Noda, et al, Effect of cell culture density on dental pulp-derived mesenchymal stem cells with reference to osteogenic differentiation. *Scientific Reports,* 2019, 9.1:5430 [MS-9096U, M or V]
https://www.nature.com/articles/s41598-019-41741-w.pdf

7. S. P. Grogan, et al, Cartilage tissue engineering combining microspheroid building blocks and microneedle arrays. *Connective Tissue Research,* 2019, 61(2) [MS-9096U]
https://www.tandfonline.com/doi/full/10.1080/03008207.2019.1617280?scroll=top&needAccess=true

< Vascular Research >

1. I. Pitaktong, et al, Early Vascular Cells Improve Microvascularization Within 3D Cardiac Spheroids. *Tissue Engineering Part C: Methods,* 2020, 26.2: 80-90 [MS-9096U]
https://www.liebertpub.com/doi/abs/10.1089/ten.tec.2019.0228

2. M. Itoh, et al, Development of an immunodeficient pig model allowing long-term accommodation of artificial human vascular tubes. *Nature Communications*, 2019, 10.1: 2244 [MS-9096U]
https://www.nature.com/articles/s41467-019-10107-1

3. K. Arai, et al, Cryopreservation method for spheroids and fabrication of scaffold-free tubular constructs. *PLoS ONE,* 2020, 15(4): :e0230428 [MS-9096U, V or M]
https://journals.plos.org/plosone/article?id=10.1371/journal.pone.0230428

4. Y. Isshiki, et al, Co-Culture of a Brain Organoid Derived from Human iPSCs and Vasculature on a Chip. *2020 IEEE 33rd International Conference on Micro Electro Mechanical Systems (MEMS)*, 2019, 1024-1027 [MS-9096V]

https://ieeexplore.ieee.org/document/9056422

< Infertility Research >

1. Y. Miyazaki, et al, Versican V1 in human endometrial epithelial cells promotes. *Reproduction*, 2019, 157.1: 53-64. [PrimeSurface dish/plate]
https://rep.bioscientifica.com/view/journals/rep/157/1/REP-18-0333.xml

2. T. Ishii, et al, Mild hypothermia promotes the viability of in vitro-produced bovine. *Journal of Reproduction and Development,* 2019, 65.3: 275-280. [MS-9096U]
https://www.jstage.jst.go.jp/article/jrd/65/3/65_2018-142/_article

3. E. S. Choi, et al, Effects of pyruvate and dimethyl-α-ketoglutarate, either alone. *Reproductive Medicine and Biology*, 2019, 18.4: 405-410. [MS-9096U]
https://onlinelibrary.wiley.com/doi/10.1002/rmb2.12288

< Immunotherapy >

1. N. Hiramatsu, et al, An analysis of monocytes and dendritic cells differentiated from human peripheral blood monocyte-derived induced pluripotent stem cells. *Medical Molecular Morphology*, 2020, 53: 63-72. [PrimeSurface dish/plate]
https://link.springer.com/article/10.1007/s00795-019-00231-8

< Transplantation and Cellular therapy >

1. H. Ueyama, et al, *S*Local transplantation of adipose-derived stem cells has a significant therapeutic effect in a mouse model of rheumatoid arthritis*cientific Reports,* 2020, 10.1: 3076 [MS-9096V]
https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7033196/pdf/41598_2020_Article_60041.pdf

< Aggregation of iPSC >

1. Y. Kato, et al, Effect of liquid flow by pipetting during medium change on deformation of hiPSC aggregates. *Regenerative Therapy,* 2019, 12: 20-26. [MS-9096V]
https://www.sciencedirect.com/science/article/pii/S2352320419300215

< Others >

1. Y, Takahashi, et al. Therapeutic potential of spheroids of stem cells from human exfoliated deciduous teeth for chronic liver fibrosis and hemophilia a. *Pediatric surgery international*, 2019, 35.12: 1379-1388. [PrimeSurface dish/plate]
https://link.springer.com/article/10.1007/s00383-019-04564-4

2. M. Oka, et al. Exogenous cytokine-free differentiation of human pluripotent stem cells into classical brown adipocytes. *Cells*, 2019, 8.4: 373. [MS-9096V]
https://www.mdpi.com/2073-4409/8/4/373

3. D. Taniguchi, et al. Human lung microvascular endothelial cells as potential alternatives to human umbilical vein endothelial cells in bio-3D-printed trachea-like structures. *Tissue and Cell*, 2020, 63:

101321. [MS-9096U]
https://www.sciencedirect.com/science/article/abs/pii/S0040816619303660

4. Y. Nakanishi, et al. Histological evaluation of tendon formation using a scaffold-free three-dimensional-bioprinted construct of human dermal fibroblasts under in vitro static tensile culture. *Regenerative therapy*, 2019, 11: 47-55. [MS-9096U, M or V]
https://www.sciencedirect.com/science/article/pii/S2352320418301007

5. Y. Takeoka, et al. Regeneration of esophagus using a scaffold-free biomimetic structure created with bio-three-dimensional printing. *PLoS ONE*, 2019, 14.3: e0211339. [MS-9096U, M or V]
https://journals.plos.org/plosone/article/file?type=printable&id=10.1371/journal.pone.0211339

6. T. Kageyama, et al. Preparation of hair beads and hair follicle germs for regenerative medicine. *Biomaterials*, 2019, 212: 55-63. [MS-90350 and MS-9096U]
https://reader.elsevier.com/reader/sd/pii/S0142961219302674?token=A5C61985425BDA2BB989D2155D089EEDA9F942639E3FAC653E381D1C02DF89ED8CED582D71F045A77E9DC2A05DF4C2A6

7. K. Suzuki, et al. Directed differentiation of human induced pluripotent stem cells into mature stratified bladder urothelium. *Scientific reports*, 2019, 9.1: 1-13. [MS-9096V]
https://www.nature.com/articles/s41598-019-46848-8

PrimeSurface® Oncology Research Publications (26)

1. M. Zanoni, et al, Anticancer drug discovery using multicellular tumor spheroid models. *Expert Opinion Drug Discovery,* 2019, 14.3: 289-301. [PrimeSurface dish/plate]
https://www.tandfonline.com/doi/abs/10.1080/17460441.2019.1570129

2. Y. Matsumoto, et al, ESTIMATION OF RBE VALUES FOR CARBON-ION BEAMS IN THE WIDE DOSE RANGE USING MULTICELLULAR SPHEROIDS. *Radiation Protection Dosimetry,* 2019, 183.1-2: 45-49. [PrimeSurface dish/plate]
https://academic.oup.com/rpd/article-abstract/183/1-2/45/5281265?redirectedFrom=fulltext

3. L. Zhao, et al, A 3D Printed Hanging Drop Dripper for Tumor Spheroids Analysis Without Recovery. *Scientific Reports,* 2019, 9.1: 19717. [MS-9096UZ]
https://www.nature.com/articles/s41598-019-56241-0

4. Y. Nashimoto, et al, Vascularized cancer on a chip: The effect of perfusion on growth and drug delivery of tumor spheroid. *Biomaterials,* 2020, 229:119547. [MS-9096U]
https://www.sciencedirect.com/science/article/abs/pii/S0142961219306465?via%3Dihub

5. J. Ko, et al, Tumor spheroid-on-a-chip: a standardized microfluidic culture platform for investigating tumor angiogenesis. *Lab on a Chip,* 2019, 19.17: 2822-2833. [MS-9096UZ]
https://pubs.rsc.org/en/content/articlelanding/2019/lc/c9lc00140a

6. J. Kondo, et al, High-throughput screening in colorectal cancer tissue-originated spheroids. *Cancer Science,* 2019, 110.1: 345-355. [MS-9384U]
https://onlinelibrary.wiley.com/doi/pdf/10.1111/cas.13843?__cf_chl_jschl_tk__=lsZRnq.NGjpHDCcq

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7. S. Kaneda, et al, Boyden chamber-based compartmentalized tumor spheroid culture system to implement localized anticancer drug treatment. *Biomicrofluidics,* 2019, 13.5: 054111. [PrimeSurface dish/plate]
https://aip.scitation.org/doi/10.1063/1.5125650

8. Y. Jimma, et al, Aryl Hydrocarbon Receptor Mediates Cell Proliferation Enhanced by Benzo[a]pyrene in Human Lung Cancer 3D Spheroids. *Cancer Investigation,* 2019, 37.8: 367-375. [MS-9096V]
https://www.tandfonline.com/doi/abs/10.1080/07357907.2019.1655760?journalCode=icnv20

9. X. Y. Qin, et al, Inhibition of Stearoyl-CoA Desaturase-1 Activity Suppressed SREBP Signaling in Colon Cancer Cells and Their Spheroid Growth. *Gastrointestinal Disorders,* 2019, 1.1: 191-200. [MS-9096U]
https://www.mdpi.com/2624-5647/1/1/14?__cf_chl_managed_tk__=zpeci7aZ0Y3oeLhc1KJqq9SgGwxhsrVroggwS_.nUyU-1642470370-0-gaNycGzNCiU

10. R. Maruhash, et al, Chrysin enhances anticancer drug-induced toxicity mediated by the reduction of claudin-1 and 11 expression in a spheroid culture model of lung squamous cell carcinoma cells. *Scientific Reports,* 2019, 9.1: 13753. [MS-9096V]
https://www.nature.com/articles/s41598-019-50276-z.pdf

11. E. Svirshchevskaya, et al, Characteristics of multicellular tumor spheroids formed by pancreatic cells expressing different adhesion molecules. *Life Sciences,* 2019, 219: 343-352. [PrimeSurface dish/plate]
https://www.sciencedirect.com/science/article/abs/pii/S0024320519300487?via%3Dihub

12. W. Masatoshi, et al, Fatty Acid β-Oxidation-dependent and -independent Responses and Tumor Aggressiveness Acquired Under Mild Hypoxia. *Anticancer Research,* 2019, 39.1: 191-200. [MS-9096U]
https://ar.iiarjournals.org/content/39/1/191.short

13. L. Houdaihed, et al, Dual-Targeted Delivery of Nanoparticles Encapsulating Paclitaxel and Everolimus: a Novel Strategy to Overcome Breast Cancer Receptor Heterogeneity. *Pharmaceutical Research*, 2020, 37.3: 39 [MS-9096U]
https://link.springer.com/article/10.1007/s11095-019-2684-6

14. K. Matsumoto, et al, Destruction of tumor mass by gadolinium-loaded nanoparticles irradiated with monochromatic X-rays: Implications for the Auger therapy. *Scientific Reports,* 2019, 9.1: 13275. [MS-9096U]
https://www.nature.com/articles/s41598-019-49978-1

15. H. Nasako, et al, Claudin-2 binding peptides, VPDSM and DSMKF, down-regulate claudin-2 expression and anticancer resistance in human lung adenocarcinoma A549 cells. *Biochimica et Biophysica Acta - Molecular Cell Research,* 2020, 1867.4: 118642. [MS-9096U]
https://www.sciencedirect.com/science/article/pii/S0167488919302502

16. R. Asai, et al, CD44 standard isoform is involved in maintenance of cancer stem cells of a hepatocellular carcinoma cell line. *Cancer Medicine,* 2019, 8.2: 773–782. [MS-9090X PrimeSurface 90mm Dish]

https://onlinelibrary.wiley.com/doi/full/10.1002/cam4.1968

17. T Futami, et al, Identification of a novel oncogenic mutation of FGFR4 in gastric cancer. *Scientific Reports,* 2019, 9.1:14627. [MS-9096U, M or V]
https://www.nature.com/articles/s41598-019-51217-6.pdf?origin=ppub

18. X. Y. Qin, et al, Lipid desaturation-associated endoplasmic reticulum stress regulates MYCN gene expression in hepatocellular carcinoma cells. Cell Death & Disease, 2020, 11: 66. [MS-9096U]
https://www.nature.com/articles/s41419-020-2257-y#citeas

19. N. X. D. Mai, et al, Biodegradable Periodic Mesoporous Organosilica (BPMO) Loaded with Daunorubicin: A Promising Nanoparticle-Based Anticancer Drug. *ChemMedChem.* 2020, 15.7: 593–599. [MS-9096U]
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20. Y. Tambe, et al, Antitumor activity of potent pyruvate dehydrogenase kinase 4 inhibitors from plants in pancreatic cancer. *Molecular Carcinogenesis,* 2019, 58.10:1726-1737. [MS-9096V]
https://onlinelibrary.wiley.com/doi/abs/10.1002/mc.23045

21. Y. Maru, et al, Establishment and characterization of patient-derived organoids from a young patient with cervical clear cell carcinoma. *Cancer Science,* 2019, 110.9: 2992-3005. [MS-9096U]
https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6726688/

22. M. M. Lübtow, et al, In vitro blood-brain-barrier permeability and cytotoxicity of atorvastatin-loaded nanoformulation against glioblastoma in 2D and 3D models. *Molecular Pharmaceutics,* 2020, 17.6: 1835-1847. [MS-9096W]
https://s3-eu-west-1.amazonaws.com/itempdf74155353254prod/10067993/In_Vitro_Blood-Brain-Barrier_Permeability_and_Cytotoxicity_of_Atorvastatin-Loaded_Nanoformulation_Against_Glioblastoma_i_v1.pdf

23. H. J. Ahn, et al, Radiation-Induced CXCL12 Upregulation via Histone Modification at the Promoter in the Tumor Microenvironment of Hepatocellular Carcinoma. *Molecules and Cells.* 2019, 42.7: 530–545. [MS-90350]
https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6681868/

24. M. Kitazawa, et al, Promotion of the Warburg effect is associated with poor benefit from adjuvant chemotherapy in colorectal cancer. *Cancer Science,* 2020, 111.2: 658-666. [MS-9096U, M or V]
https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7004516/

25. R. Kikuchi, et al, Hypercapnic tumor microenvironment confers chemoresistance to lung cancer cells by reprogramming mitochondrial metabolism in vitro. *Free Radical Biology and Medicine,* 2019, 134: 200-214. [MS-9096U]
https://www.sciencedirect.com/science/article/abs/pii/S0891584918313030

26. T, Sulea, et al, Structure-based engineering of pH-dependent antibody binding for selective targeting of solid-tumor microenvironment. *MAbs,* 2020, 12.1: 1682866. [MS-9096U, M or V]
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PrimeSurface® Other Publications (11)

1. Y. Inubushi, et al, Uniform spheroid formation on a laboratory-made, low cell attachment surface consisting of a chitin sheet. *Biochemistry & Molecular Biology,* 2019, 84.5: 997-1000. [MS-9096U, M or V]
   https://www.tandfonline.com/doi/full/10.1080/09168451.2020.1714423

2. S. Kozaki, et al, Additive Manufacturing of Micromanipulator Mounted on a Glass Capillary for Biological Applications. *Micromachines*, 2019, 11.2: 174. [MS-9096U]
   https://www.mdpi.com/2072-666X/11/2/174

3. K. Iuchi, et al, Different morphologies of human embryonic kidney 293T cells in various types of culture dishes. *Cytotechnology,* 2019, 72: 131-140. [PrimeSurface dish/plate]
   https://link.springer.com/article/10.1007/s10616-019-00363-w

4. K. Sugihara, et al. Mechanisms of endothelial cell coverage by pericytes: computational modelling of cell wrapping and in vitro experiments. *Journal of the Royal Society Interface*, 2020, 17.162: 20190739. [MS-9096U]
   https://royalsocietypublishing.org/doi/full/10.1098/rsif.2019.0739

5. S. Floriana, et al, A human organoid system that self-organizes to recapitulate growth and differentiation of a benign mammary tumor. *PNAS,* 2019, 116.23: 11444-11453. [MS-9096U]
   https://www.pnas.org/content/116/23/11444

6. T. Kazi et al, Optimal stimulation toward the dermal papilla lineage can be promoted by combined use of osteogenic and adipogenic inducers. *FEBS*, 2020, 10.2: 197-210. [MS-9096U, M or V]
   https://febs.onlinelibrary.wiley.com/doi/full/10.1002/2211-5463.12763

7. W. K. Restu et al, Hydrogel formation by short D-peptide for cell-culture scaffolds. *Materials Science and Engineering: C*, 2020, 111: 110746. [PrimeSurface dish/plate]
   https://www.sciencedirect.com/science/article/abs/pii/S0928493119331856

8. M. Mehruba et al, PKN1 controls the aggregation, spheroid formation, and viability of mouse embryonic fibroblasts in suspension culture. *Biochemical and Biophysical Research Communications*, 2020, 523.2: 398-404. [PrimeSurface dish/plate]
   https://www.sciencedirect.com/science/article/abs/pii/S0006291X19323976

9. C. S. Ong et al, Principles of Spheroid Preparation for Creation of 3D Cardiac Tissue Using Biomaterial-Free Bioprinting. *3D Bioprinting*, 2020, 2140: 183-197. [PrimeSurface dish/plate]
   https://link.springer.com/protocol/10.1007/978-1-0716-0520-2_12

10. P. Zhang et al, Knockdown of long non-coding RNA PCAT1 in glioma stem cells promotes radiation sensitivity. *Medical Molecular Morphology*, 2019, 52: 114-122. [PrimeSurface dish/plate]
    https://link.springer.com/article/10.1007/s00795-018-0209-8

11. T. Inagaki et al, Direct evidence of abortive lytic infection-mediated establishment of Epstein-Barr virus latency during B-cell infection. *Frontiers in Microbiology*, 2020, 11: 575225. [MS-90350]
https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7888302/

Publications in 2018 (46)

1. K. Tsuji-Tamura, et al, Dual inhibition of mTORC1 and mTORC2 perturbs cytoskeletal organization and impairs endothelial cell elongation. *Biochemical and Biophysical Research Communications*, 2018, 497.1.:326-331 [MS-9096U]
https://www.sciencedirect.com/science/article/pii/S0006291X18303036

2. R. Akizukia, et al, Decrease in paracellular permeability and chemosensitivity to doxorubicin by claudin-1 in spheroid culture models of human lung adenocarcinoma A549 cells. Biochimica et Biophysica Acta (BBA) - *Molecular Cell Research*, 2018, 1865.5.:769-780 [MS-9096U]
https://www.sciencedirect.com/science/article/pii/S0167488918300363

3. E. C. Costa, et al, Spheroids Formation on Non-Adhesive Surfaces by Liquid Overlay Technique: Considerations and Practical Approaches. *Biotechnology Journal (Special Issue: Biotech Methods and Advances)*, 2018, 13.1 [MS-9096U]
https://onlinelibrary.wiley.com/doi/abs/10.1002/biot.201700417

4. Y Fukuda-Takami, et al, Layer-by-layer cell coating technique using extracellular matrix facilitates rapid fabrication and function of pancreatic β-cell spheroids. *Biomaterials.* 2018, 160.:82-91 [MS-9096U]
https://www.sciencedirect.com/science/article/pii/S0142961218300267

5. R. Maruhashia, et al, Elevation of sensitivity to anticancer agents of human lung adenocarcinoma A549 cells by knockdown of claudin-2 expression in monolayer and spheroid culture models. Biochimica et Biophysica Acta (BBA) - *Molecular Cell Research, 2018*, 1865.3.:470-479 [MS-9096U]
https://www.sciencedirect.com/science/article/pii/S0167488917303208

6. J. Terashima, The regulation mechanism of AhR activated by benzo[a]pyrene for CYP expression are different between 2D and 3D culture of human lung cancer cells. *Drug Metabolism and Pharmacokinetics*, 2018 [MS-9096V]
https://www.sciencedirect.com/science/article/pii/S1347436717302318

7. H. Yamamoto, et al, Characterization of genetically engineered mouse hepatoma cells with inducible liver functions by overexpression of liver-enriched transcription factors. *Journal of Bioscience and Bioengineering*, 2018, 125.1.:131-139 [MS-9096 U, M or V]
https://www.sciencedirect.com/science/article/pii/S138917231730556X

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